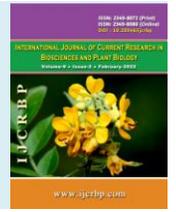




International Journal of Current Research in Biosciences and Plant Biology

Volume 9 • Number 2 (February-2022) • ISSN: 2349-8080 (Online)

Journal homepage: www.ijcrbp.com



Review Article

doi: <https://doi.org/10.20546/ijcrbp.2022.902.004>

Bovine *in vitro* embryo production and its contribution in improving animal products and productivity: A review

Mosisa Dire Babura*

Ethiopian Institute of Agricultural Research, Debre Zeit Agricultural Research Center,
P.O. Box 32, Debre Zeit- Ethiopia

*Corresponding author; e-mail: mosisamd43@gmail.com

Article Info

Abstract

Keywords:

In vitro culture
Embryo production
Fertilization
Media
Oocyte

In vitro embryo production of bovine embryos has become implemented in cattle breeding in many countries for the production of embryos in the laboratory which has several advantages as compared with other conventional methods of embryo production. The various aspects of In-vitro maturation, fertilization, and embryo culture in domestic farm animals are the major components of IVEP where to be considered due to *In vitro* embryo development being strongly influenced by events occurring during these steps of the techniques. Many factors can affect the success of IVEP by affecting all the steps of the techniques or at least one step. Therefore this paper was reviewed for the objective of considering the factors that influence the efficiency of *In vitro* embryo production. So, for improving the efficiency of IVP and making the systems more important for routinely producing blastocyst stages of embryos for high genetic merit, factors affecting *In vitro* maturation, fertilization, and embryo culture should be considered.

• Received: 23 November 2021 • Revised: 15 January 2022 • Accepted: 24 January 2022 • Published Online: 6 February 2022

Introduction

The world food crisis and rise in demand for food of animal origin, competitive global markets of such foods, and the negative effects of climatic change, especially in the marginal tropical areas (Thornton et al., 2006), demand that innovative strategies and interventions be urgently employed (Sere et al., 2008). The dairy sector in developing countries, especially in low-input production systems, has the potential of providing cost-effective animal protein requirements. In order to provide high-quality meat and dairy products, the enhancement of domestic production is the most

permanent method. Thus, the implementation of advanced animal improvement programs is essential to make animal agriculture sustainable and more profitable. Overall, advanced animal biotechnologies are essential tools to accelerate the improvement of genetics and allow the selection of high-producing, fertile, and healthy animals in developing countries. These reproductive biotechnologies include artificial insemination (AI), semen technologies and evaluation, embryo transfer, *in vitro* fertilization (IVF), ovum pickup and combination with IVF, estrus synchronization, superovulation, and pregnancy monitoring (Kaya et al., 2018).

Over the past years, *in vitro* production (IVP) of bovine embryos has become implemented in cattle breeding in many countries. Compared with conventional superovulation and ET, the production of embryos in the laboratory has several advantages. First, IVEP can be used in donors those fail to respond to superovulation treatment. Second, IVEP can be used to salvage the genetic potential of terminally ill females that could not respond to conventional ET. Third, semen from different bulls can be used to fertilize oocytes harvested from a cow resulting in embryos with different sires being produced at the same time. Fourth, oocytes for IVEP can be obtained from the ovaries of the live donors using Transvaginal Oocyte Recovery (TVOR), or from the slaughter ovaries (Suthar and Shah, 2009).

The development of techniques for the effective production of bovine preimplantation embryos from oocytes matured and fertilized *in vitro* is important for embryo transfer and basic scientific research. Techniques for producing pre-implantation embryos by *in vitro* maturation/*in vitro* fertilization (IVM/IVF) are being used in many laboratories worldwide (Ocana Queo et al., 1999).

In the laboratory, embryos can be routinely produced and developed up to the blastocyst stage using three subsequent techniques: *In vitro* maturation (IVM) of oocytes, followed by sperm capacitation and *in vitro* fertilization (IVF) of matured oocytes and then *in vitro* culture (IVC) of the fertilized oocytes up to the blastocyst stage. *In vitro* embryo development is strongly influenced by events occurring during oocyte maturation, fertilization, and the subsequent development of the fertilized oocytes. So, improving the efficiency and identifying the sources of variations between IVF systems are more important for routinely producing blastocyst stages of embryos for high genetic merit (Kaya et al., 2018).

Therefore the objectives of this review are:

- To review the optimal condition required for IVM, IVF, IVC, and factors affecting their success
- To review the different semen preparation techniques and their effects on In-vitro fertilization of the matured oocyte
- To review the types of media used in each step of IVEP

***In vitro* maturation**

Oocyte maturation is a long process during which oocytes acquire their intrinsic ability to support the subsequent stages of development in a stepwise manner, ultimately reaching activation of the embryonic genome. This process involves complex and distinct, although linked, events of nuclear and cytoplasmic maturation. Nuclear maturation mainly involves chromosomal segregation, whereas cytoplasmic maturation involves organelle reorganization and storage of mRNA, proteins, and transcription factors that act in the overall maturation process, fertilization, and early embryogenesis (Motlagh et al., 2008). The maturation of the oocytes includes two aspects: Nuclear and cytoplasmic maturation (Roa et al., 2002).

Oocyte selection for *in vitro* maturation

For the most successful species, it was only possible to obtain the development of a maximum of 40 to 50% of the zygotes to the blastocyst stage (Watson 2007). This limited success had been attributed to: the reduced oocyte developmental competence which was suggested to be the primary reason for the reduced potential of *in vitro* produced embryos or due to the heterogeneous population of oocytes obtained from the follicles (Sirard et al., 2006; Vanblerkam 1990).

Bovine oocyte selection for IVM was performed on the basis of visual assessment for the morphological features (Leibfried, 1979). Cumulus cells establish intimate contacts together as well as with the oocyte to allow cell communication through gap junctions, which were necessary for efficient maturation of the oocyte (Vansoom et al., 2002).

The criteria for oocyte selection depend not only on the morphology of the oocyte-cumulus complex but also upon other factors such as the follicular size (Klumpp 2004). Where, as a lot of researches show that the developmental capacity of the bovine oocyte is affected by the follicular size (Carolan et al., 1996). The capacity of the bovine oocyte derived from different follicular sizes to undergo normal fertilization and early embryonic development *in vitro* and reported that the follicles could categorize into three groups: > 4 to 8 mm (large), > 2 to 4 mm (medium), and 2 to 1mm in diameter (small) were, there were high percentages of embryonic development and blastocyst rate recovered from the large follicles (Pavlok et al., 1992). Similarly,

(Mate and Rodgers, 1993) stated that there was an effect for the follicular size on the oocyte developmental competence where the percentage of the oocytes reached M-II from large follicles (=1.5 mm) was 55% compared to 20% recovered from small follicles (<1.5 mm). (Lonergan et al., 1994) added that the oocytes obtained from 6-7mm follicles usually had many layers of cumulus cells and significantly produced a higher proportion of morula/blastocyst stage embryos (65.9%) compared to those obtained from 2-6 mm follicles (34.3%).

Overall the morphological criteria are still routinely used for a rough selection of the oocytes in most laboratories as there was no real breakthrough with respect to increased blastocyst rates that had been achieved yet (Hyttel et al., 1997).

Factors affecting *in-vitro* maturation

In vitro maturation is the most critical step *in vitro* embryo production. There is a constant need to emphasize the fact that effective oocyte maturation is the foundation of embryo production. Identifying these factors will improve the *in vitro* embryo production systems in bovine (Kaya et al., 2018). The recovery of a large number of oocytes with high developmental competence remains an ultimate goal for the mass production of embryos in cattle. At the same time, the origin of the oocyte can play an important role in their IVF and subsequent developmental competence.

Effects follicular size

Oocytes for IVM are generally selected using the following criteria: follicle size, cytoplasmic appearance, the appearance, and a number of cumulus cells around the oocytes (COCs). Cumulus expansion can importantly be used to microscopically assess the *in vitro* maturation rate of oocytes and also showed a relationship between follicle size and oocyte quality (Gubta et al., 2005; Lonergan et al., 1994a; Yang et al., 1998).

In bovine, the oocyte first acquired competence to develop into blastocysts *in vitro* system at a follicular size of 2-3 mm. When follicles were pooled according to size, it was shown that large follicles (10 mm diameter) contain oocytes with a higher potential to become embryos (Lonergan et al., 1994). Some studies described the fate of individual oocytes according to the

exact follicular size and it confirmed an increased competence with follicle size, i.e., bovine oocyte complexes (COCs) isolated from ovaries carrying follicles of 2-5 mm in diameter showed lower rates of maturation and blastocyst formation than those from ovaries carrying follicles of >10 mm in diameter (Kubota and Yang 1998). These indicate that large follicles (6 mm diameter) provide the oocyte with a microenvironment that improves its quality (Lonergan, 1992).

It has been recorded that the follicular size was not the only criterion determining the oocyte competence to develop, as there were some oocytes originating from large follicles failed to produce embryos, while others from small ones already have this capacity; this simply because the oocyte capacity to mature, fertilize and to develop into blastocysts was acquired in a stepwise fashion during oogenesis and folliculogenesis (Eppig, 1996).

The existence of cumulus cells

The presence of cumulus cells was necessary for the cytoplasmic and /or nuclear maturation of oocytes in cattle (Zhang et al., 1995). Cumulus cells benefit oocytes development either by secreting soluble factors, which induced developmental competence or by removing an embryo development suppressive component from the medium (Hashimoto et al., 1998). Cumulus cells supported the IVM of oocytes to the MII stage and were involved in the cytoplasmic maturation needed for optimal developmental competence, such as male pronucleus formation and development to the blastocyst stage. Cumulus cells might be a good indicator for an oocyte's ability to undergo meiosis I *in vitro* and that the developmental problems of denuded oocytes were due to deficient cytoplasmic maturation (Davachi et al., 2012).

Various roles of cumulus cells include prevention of the hardening of zona pellucida, the provision of energy for oocyte maturation and production of cytoplasmic maturation factors, and the uptake of nutrients for oocytes during maturation in culture medium (Mori et al., 2000). In addition, the cumulus cells are also important for fertilization, for example, such as the trapping of spermatozoa, guiding spermatozoa to the oocyte, protection of the oocytes against zona hardening, and prevention of changes in the oocyte that are unfavourable for subsequent fertilization (Vansoom

et al., 2002; Tanghe et al., 2002). Moreover, factors like chemokines secreted from COCs induce sperm capacitation and enhance fertilization, providing evidence for a regulatory loop between sperm and COCs during fertilization (Shimada et al., 2008). It has been demonstrated that chemokine signalling facilitates both sperm attraction to the COC and COC compaction by the cumulus extracellular matrix assembly (Tamba et al., 2008).

Oocyte quality

Naturally, the oocyte quality is determined by the oocyte's ability to mature, be fertilized, and give rise to normal offspring (Duranton and Renard, 2001; Hussein et al., 2006). The quality of the oocyte is also related to the oocytes' follicular environment, as well as several factors: such as the age of the donor animal, stage of follicular development, and the media used for maturing the oocytes (Camargo et al., 2006; Keskintepe et al., 1994).

Cumulus cells morphology and the microscopic aspect of the ooplasm are generally considered as the two main parameters to assess the quality of the cumulus-oocyte complex (COC) (Haseleger et al., 1995). The criteria employed by various authors for the selection of oocytes for IVM include the presence of a multilayer compact cumulus oophorus and homogeneous cytoplasm (De Wit et al., 2000).

Effect of culture media

A wide variety of media has been used for IVM in domestic animals, ranging from simple physiological solutions to complex culture media containing amino acids, vitamins, purines, and other compounds regarded as essential for general cell culture. The culture employed in IVM not only affects the proportion of bovine oocytes that reach metaphase II (M II) and become capable of undergoing *in vitro* fertilization but can also influence subsequent embryonic development (Bavister, 1992).

In vitro maturation medium can be broadly divided into simple and complex. Simple media are usually bicarbonate-buffered systems containing physiological saline with pyruvate, lactate, and glucose and they differ in their ion concentration and in the concentrations of the energy sources. Complex media contains in addition to the basic components of simple media, amino acids,

vitamins, and purines.

Most IVF laboratories routinely use M-199 as the basic IVM medium in cattle and there have been few reports suggesting that other media may be more appropriate. In one comparison of complex media for IVM, the scientists concluded that under their conditions, the F-10 medium is superior to M-199 and B2 media (Hawk and wall, 1993). Another comparison of IVM commercially available complex chemically defined media showed that TCM-199 was superior to RPMI-1640 (Gliedt et al., 1996). Oocytes matured in medium leading to poor developmental competence have depressed levels of glycolysis that are necessary for the completion of maturation, the reduced level of glycolysis may reflect the reduced activity of the pentose phosphate pathways, which plays an important role in the meiotic maturation of bovine oocytes (Krisher and Bavister, 1998). For the energy source, the excessive glucose in the media used for oocyte maturation impairs the development of bovine oocytes to the blastocyst stage, possibly due to the increase of Reactive Oxygen Species (ROS) and the decrease in the intracellular glutathione content of bovine oocytes (Hashimoto et al., 2000).

In vitro fertilization

The IVF is the mixed cultivation of mature oocytes with spermatozoa in strictly controlled laboratory conditions. The co-incubation of spermatozoa with oocytes lasts for 24 hours, at 39 °C with 5% CO₂ in the air and 80% humidity after which the first mitosis occurs (Lojkic et al., 2014). The success of IVF of bovine oocytes is estimated 48 hours following fertilization by a number of cleaved embryos and by identification of male and female pronucleus which may be visible after 18-22 hours of co-incubation (Leibfreid et al., 1996). Co-incubations are usually conducted in 50 to 100 fill micro droplets with 5 to 25 oocytes/drop-in Petri dishes covered with mineral oil. The sperm concentration for *in vitro* fertilization ranges from 1 to 20 x 10⁶ motile sperm/ml of medium and oocytes are usually co-incubated with sperm for 6 to 24 hours. Finally, it was washed several times to remove extra sperm cells and then transferred into embryo development culture medium (Goto et al., 1989).

Factors affecting *in vitro* fertilization

Fertilization is a complex process, which results in the

union of two gametes, the restoration of the somatic chromosome number, and the start of the development of a new individual. Successful cattle IVF requires appropriate preparation of both sperm and oocyte, as well as culture conditions that are favorable to the metabolic activity of the male and female gametes (Xu and King 1990).

Methods of sperm preparation

Mammalian spermatozoa, matured in the epididymis and ejaculated, are not immediately capable of fertilizing oocytes. The sperm must undergo a period of capacitation, which normally occurs in the female reproductive tract.

Preparing sperm for fertilization of the bovine oocyte involves a sequence of events in which the sperm: One is motile (to reach the oocyte and move through the Zona Pellucida (ZP), two the ability to undergo capacitation and express the Acrosome Reaction (AR), Third is the capacity to bind to the zona pellucida and vitelline membrane by acquiring the correct binding proteins during maturation and exposing these binding sites to the oocyte at the appropriate time and fourth is the ability to fuse with the oolemma and be incorporated into the oocyte.

It is clearly important to have highly motile bull sperm available for IVF. This may be achieved by applying various procedures for isolating motile samples. There are also a number of chemical agents which may be employed to stimulate motility and AR of bull sperm and to maintain motility (Kaya et al., 2018).

Percoll density gradient method

In the procedure of this method 1.5 ml of 90% Percoll was mixed with 1.5 ml of sperm- Tyrode's-Lactate (Sp-TL) stock to obtain 3 ml of 45% Percoll in a 15 ml tube. With a transfer pipette, 90% Percoll (3 ml) was carefully deposited under 45% Percoll in the bottom of the tube. Semen (0.25 ml) was deposited on the top of the Percoll gradient.

This was centrifuged at 400G for 30 minutes at room temperature (25 °C). A sperm pellet formed at the bottom was removed and mixed with Sp-TL to make a final volume of 5 ml which was again centrifuged for 10 minutes. After discarding supernatant, the pellet was examined for sperm concentration and diluted with

Tyrodealbumin- lactate-pyruvate (TALP: NaCl 99.0 mM, KCl 3.1 mM, NaHCO₃ 25.0 mM, NaH₂PO₄ 0.35 mM, bovine serum albumin 6.0 mg/ml, sodium pyruvate 1.0 mM, DL-lactic acid 21.6 mM, Hepes 10 mM) medium to have final concentration of 1x10⁶/ml of TALP (Rosenkrans et al., (1993).

Swim-up method

Swim-up method was performed with two types of media separately; i) Modified Ca²⁺ free Tyrode's medium and ii) Tyrodealbumin- lactate-pyruvate (TALP) medium. Each medium (pH 7.3-7.4) was incubated in an atmosphere of 5% CO₂ in air at 38.5 °C for 2 hours prior to use. A 0.25 ml of fresh semen was deposited at the bottom of 1.5 ml of each medium separately. The tubes were incubated in an atmosphere of 5% CO₂ in air at 38.5°C for 30 minutes and supernatant from the same media tubes was pooled separately in a sterile conical tube and centrifuged at 100G for 10 minutes. The supernatant was discarded, saving 100 µl sperm suspension at the bottom of each tube. This sperm suspension in each tube was diluted with 1 ml of the respective medium. This preparation was equilibrated at room temperature for 5 minutes. After adding 5 ml of more medium, it was again centrifuged for 10 minutes at 100 G. The supernatant was again discarded, and the remaining 100 µl of sperm suspension in each tube was diluted with the same medium containing heparin (21.87 IU/ml) and incubated finally for 15 minutes in CO₂ incubator at 38.5°C (Parrish et al., 1985).

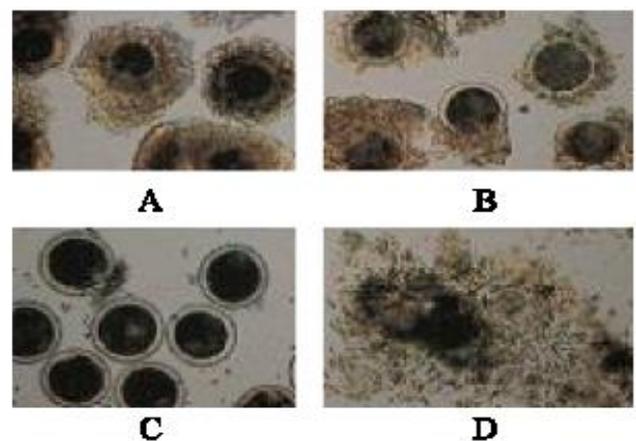


Fig. 1: Representative photographs of the four grades of COCs. Grade A- homogenous COCs, Grade- B: COCs not homogenous, Grade- C: COCs were not found at all, and Grade- D: expanded COCs.

Sodium citrate washing

In the sodium citrate washing method, 0.25 ml of fresh semen was mixed with 2.9% sodium citrate to make a final volume of 5 ml. The suspension was centrifuged at 300G for 10 minutes. The supernatant was discarded and the pellet containing sperms was dissolved in 3 ml of 2.9% sodium citrate and centrifuged again for 10 minutes.

After discarding the supernatant, the concentration was determined in the sperm pellet and sufficient Tyrode solution was added to obtain a final sperm concentration of 1×10^6 /ml of medium (Samad et al., 1998).

In vitro embryo culture and its development

The fertilized oocytes are cultured *in vitro* to the blastocyst stage when such embryos may be transferred into a recipient or frozen and stored in liquid nitrogen. Development to the morula and blastocyst was found to be dependent on the gas phase environment during *in vitro* culture. The best gas-phase observed was 5% CO₂, 5% O₂, and 90% N₂.

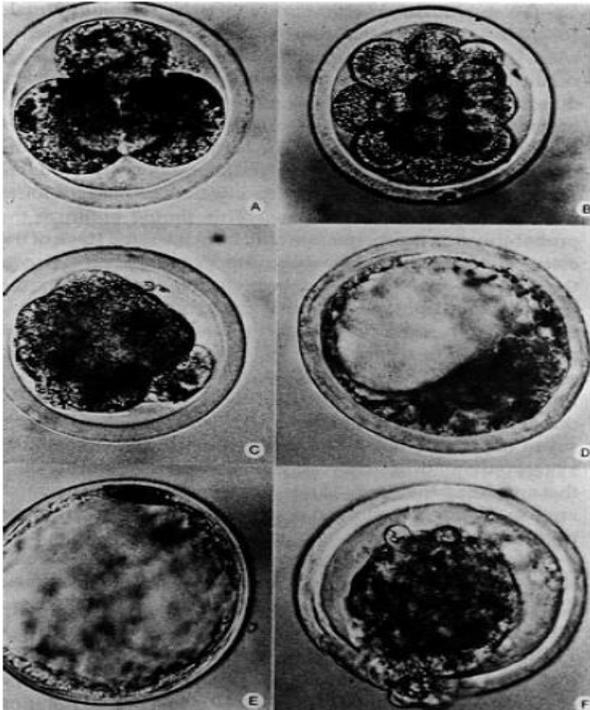


Fig. 2: Cattle embryos at various stages of development (Betteridge, 1977). A. 4-cell egg, day 3. B. 16-cell egg, day 5. C. Morula, day 6. Cells have compacted and lost individual outlines. D. Early blastocyst, day 7. E. Blastocyst fully expanded within the zona pellucida, day 10. F. Hatching blastocyst, day 10.

The IVC lasts for 7 days in desirable defined sequential media to satisfy the energy needs of the pre-implantation embryo (Samardzija et al., 2005). During IVC, the embryo is passing through 4 important developmental stages: the first mitosis, activation of the embryonic genome, compaction in morula stage, the formation of the blastocyst with differentiation of their cells. Unfavourable conditions during IVC may affect any of the mentioned stages and negatively influence the quality of cultivated blastocyst. The quality of the embryo has further influenced the success of freezing as well as the percentage of conception after transferring to the recipient animal. Although numerous methods are used to evaluate embryo quality, the assessment of this quality presents a challenge for embryologists in selecting the best quality embryos to transfer to the recipient. The simplest and most widespread method to determine the embryo quality is a morphological evaluation, which is the most subjective method (Lojkić et al., 2014).

Mammalian embryos have been cultured in a variety of chemically defined and undefined media. A chemically defined medium has been described as a liquid prepared from bench chemicals and containing four or less basic components: inorganic salts, amino acids, vitamins, and an energy source at known concentrations for lists of ingredients of several common defined media (Biggers, 1987).

Obviously, the study continued to improve methods of maturation and fertilization but embryo culture stood as the final great frontier to *in vitro* embryo production.

The benefits offered by IVC are easy to see, such as the ability to observe development in real-time, as well as the ability to non-surgically transfer embryos at a later stage of development. The latter is absolutely essential for the widespread commercial application of *in vitro* embryo production.

Although the media choices of the initial attempts at *in vitro* culture were logical as they supplied protein, nutrients, and growth factors, embryo culture in them was relatively unrewarding. Many researchers set out to find exactly what type of fluid embryos were cultured *in vivo*. The first scientists elucidated the ionic and protein contents of the oviductal fluid of sheep. Appropriately, the resulting media was called Synthetic Oviductal Fluid (SOF) and forms the basis for many ruminants' *in vitro* production protocols of today (Restall and Wales

1966). The SOF was used for the first time in bovine embryo culture (Tervit et al., 1972). The *in vivo* produced embryos that started at the eight-cell stage proceeded to the morula stage in 30% of embryos, which represented only the second time that a morula had been produced in culture. In the same study, oxygen tension was varied to better approximate the conditions of the oviduct (Whitten 1971). When O₂ levels were reduced to 5% (from the 21% found in the atmosphere) with 5% carbon dioxide and the balance nitrogen, the development of 8 cells *in vivo* produced embryos proceeded appropriately and the world saw for the first time a bovine blastocyst as the result of *in vitro* culture. The lower oxygen tension is thought to reduce the load of reactive oxygen species present in the media (Noda et al., 1991).

Early successes in bovine embryo development were often seen either from 1-2 cell embryos to the 8 cell stage or from the eight-cell stage to morula/blastocyst. Thus, the term “8 cell block” was coined to describe the insufficiency of culture systems of the day to support development through the fourth cell cycle. Interestingly, this “Block” occurs at the same point as Embryonic Genome Activation (EGA) or the shift in the embryo from the embryo’s dependence on maternal mRNA to embryonic. The application of low oxygen tension in bovine embryo culture was the first step in solving the 8-cell block (Tervit et al., 1972). The embryos in the third and fourth cell cycle were most affected by *in vitro* culture conditions and would not proceed in development *in vivo* after *in vitro* culture for 24 h, even though embryos cultured for short periods *in vitro* at other points in development could proceed to the blastocyst stage after *in vivo* culture. In addition to low oxygen tension, somatic cell co-culture has also been shown to overcome this “Block” (Eyestone and First, 1989).

Assessment of *in vitro* produced embryo quality

There are several ways to objectively measure the health and quality of *in vitro* produced embryos. The most obvious measure is to transfer them to synchronized recipients and assess pregnancy and calving rate compared to *in vivo* produced embryos transferred to another group. Indeed, although this might serve as the “Gold standard” test of embryo health, it is often not feasible or practical due to using oocytes of unknown genetic quality or financial constraints of research. There is a validated method of evaluating *in vivo* produced embryos which were

developed by members of the International Embryo Transfer Society. These guidelines concern the percentage of cellular material which forms the embryonic cell mass as well as the stage of development compared to the expected stage. They serve to inform decisions about transferring, freezing, and discarding *in vivo* produced embryos (Stringfellow and Seidel, 1998).

Important considerations for IVF

The cleanliness of the glassware is of utmost importance. Used glassware is rinsed thoroughly with tap water and soaked in biological detergent overnight. The following day, glassware is rinsed thoroughly at least 20 - 40 times in tap water, and then submerged in a 3-5% HCl for a minimum of 2 hours. It is then rinsed in running water for about 15 minutes and rinsed again 20 - 40 times in tap water. Thereafter, glassware is rinsed with distilled water and placed upside-down to dry by heating it at 120°C for 2 - 4 hours. Glassware is then packed separately in aluminium foil and sterilized by heating at 120°C for 4 hours. All ultrasonic cleaner is also used for heavily soiled and difficult to wash glassware such as volumetric flask, test tube, Pasteur, and volumetric pipettes. Cleaning by sonication takes around 15 minutes in hot water. After sonication, glassware is rinsed 20 - 40 times with tap water and finally rinsed with distilled water. Thereafter, the drying and sterilization protocol described above is practiced.

Conclusions and recommendation

The development of techniques for the effective production of bovine preimplantation embryos from oocytes matured and fertilized *in vitro* is important for embryo transfer. However *In vitro* embryo development is strongly influenced by events occurring during the three subsequent steps of the techniques: oocyte maturation, fertilization and development of fertilized oocytes. Therefore based on the above conclusion the following recommendations are forwarded: (i) Factors affecting oocyte maturation should be considered before starting maturation of with recommended quality of oocytes; (ii) Appropriate semen preparation methods should be used for *in vitro* fertilization; (iii) The optimal condition required for embryo culturing should be considered to minimize loss of fertilized zygote like that of eight cell block. The types of media and its composition or ingredients used for each steps should be considered with great attention for the amounts of ingredients.

Conflicts of Interest

There is no conflict of interest.

References

- Ayman H. Abd El-Aziz, Usama E. Mahrous, Sherief Z. Kamel and Ahmed A. Sabek, 2016. Factors influencing *in vitro* production of bovine embryos: A review. *Asian J. Anim. Vet. Adv.*, 11: 737-756.
- Bavister, B. D., T. A. Rose-Hellekant and T. Pinyopummintr, 1992. Development of *in vitro* matured/*in vitro* fertilized bovine embryos into morulae and blastocysts in defined culture media. *Theriogenology*, 37: 127-146.
- Camargo, L. S. A., J. H. M. Viana, W. F. Sa, A. M. Ferreira, A. A. Ramos and V. R. Vale Filho, 2006. Factors influencing *in vitro* embryo production. *Anim. Reprod.*, 3: 19-28.
- Carolan, C., P. Lonergan, P. Monget, D. Monniaux and P. Mermillod, 1996. Effect of follicle size and quality on the ability of follicular fluid to support cytoplasmic maturation of bovine oocytes. *Mol. Reprod. Dev.*, 43: 477-483.
- Davachi, N. D., H. Kohram and S. Zainoaldini, 2012. Cumulus cell layers as a critical factor in meiotic competence and cumulus expansion of ovine oocytes. *Small Rumin. Res.*, 102: 37- 42.
- Duranthon, V. and J. P. Renard, 2001. The developmental competence of mammalian oocytes: A convenient but biologically fuzzy concept. *Theriogenology*, 55: 1277-1289.
- Eppig, J. J., 1996. Coordination of nuclear and cytoplasmic oocyte maturation in eutherian mammals. *Reprod. Fertil. Dev.*, 8: 485-489.
- Eyestone, W. H. and N. L. First, 1989. Co-culture of early cattle embryos to the blastocyst stage with oviducal tissue or in conditioned medium. *J. Reprod. Ferti.*, 85: 715-720.
- Gliedt, D. W., C. F. Rosenkrans Jr., R. W. Rorie and J. M. Rakes, 1996. Effects of oocyte maturation length, sperm capacitation time and heparin on bovine embryo development. *J. Dairy Sci.*, 79: 532-535.
- Goto, K., Y. Kajihara, M. Koba, S. Kosaka, Y. Nakanishi and K. Ogawa, 1989. *in vitro* fertilization and development of *in vitro* matured bovine follicular oocytes. *J. Anim. Sci.*, 67: 2181-2185.
- Gupta, P. S. P., J. P. Ravindra, V. Girishkumar, H. M. Raghu and S. Nandi, 2005. Stimulation of *in vitro* oocyte maturation with a novel peptide isolated from follicular fluid of the buffalo (*Bubalus bubalis*). *Small Rumin. Res.*, 59: 33-40.
- Hashimoto, S., K. Saeki, Y. Nagao, N. Minami, M. Yamade and K. Utsumi, 1998. Effects of cumulus cell density during *in vitro* maturation of the developmental competence of bovine oocytes. *J. Theriogenology*, 49: 1451-1463.
- Hashimoto, S., N. Minami, M. Yamada and H. Imai, 2000. Excessive concentration of glucose during *in vitro* maturation impairs the developmental competence of bovine oocytes after *in vitro* fertilization: Relevance to intracellular reactive oxygen species and glutathione contents. *Mol. Reprod. Dev.*, 520-526.
- Hawk, H. W. and R. J. Wall, 1993. Experiments to improve the yield of bovine blastocysts from *in vitro* produced oocytes. *J. Anim. Sci.*, 71: 220-220.
- Hazeleger, N. L., D. J. Hill, R. B. Stubbings and J. S. Walton, 1995 Relationship of morphology and follicular fluid environment of bovine oocytes to their developmental potential *in vitro*. *Theriogenology*, 43: 509-522.
- Hussein, T. S., J. G. Thompson and R. G. Gilchrist, the 2006. Oocyte-secreted factors enhance oocyte developmental competence. *Dev. Biol.*, 296: 514-521.
- Kaya a, GÜNEŞ E, MEMİLİ E. 2018; Application of reproductive biotechnologies for sustainable production of livestock in Turkey: A review, *Turk J Vet Anim sci*, Ro 18142:143-151, doi: 10.3906/Vet-1706-66.
- Keskintepe, L., G. M. Darwish, A. T. Kenimer and B. G. Brackett, 1994. Term development of caprine embryos derived from immature oocytes *in vitro*. *Theriogenology*, 42: 527-535.
- Klumpp, A. M., 2004. The effect of holding bovine oocytes in follicular fluid on subsequent fertilization and embryonic development. M.S. thesis, Graduate Faculty of Louisiana State Univ. and Agricultural and Mechanical College.
- Krisher, R. L. and B. D. Bavister, 1998. Responses of oocytes and embryos to culture environment. *Theriogenology*, 59: 103-114.
- Kubota, C. and X. Yang, 1998. Cytoplasmic incompetence results in poor development of bovine oocytes derived from small follicles. *Theriogenology*, 49: 183-183.
- Leibfried, L. and N. L. First 1979. Characterization of bovine follicular oocytes and their ability to mature *in vitro*. *J. Anim. Sci.*, 48(1): 76-86.
- Leibfried-Rutledge M L, Critser E S, Parrish J J, First N

- L. *In vitro* maturation and fertilization of bovine oocytes. *Theriogenology*. 1996;31(1):61-74.
- Lojkić M, Čavlek M, Bačić G, Getz I, Samardžija M, Maćešić N, et al., Morphology evaluation of bovine embryos produced *in vitro*. *Vet. stn.* 2014;45(3):187-193.
- Lonergan, P., C. Carolan and P. Mermillod, 1994a. Development of bovine embryos *in vitro* following oocyte maturation under defined conditions. *Reprod. Nutr. Dev.*, 34: 329-339.
- Lonergan, P., P. Monaghan, D. Rizos, M. P. Boland and L. Gordon, 1994. Effect of follicle size on bovine oocyte quality and developmental competence following maturation, fertilization and culture *in vitro*. *Mol. Reprod. Dev.*, 37: 48-53.
- Mori, T., T. Amano and H. Shimizu, 2000. Roles of gap junctional communication of cumulus cells in cytoplasmic maturation of porcine oocytes cultured *in vitro*. *Biol. Reprod.* 62: 913-919.
- Motlagh, M. K., A. Z. Shahneh, M. Daliri, H. Kohram and F. Gharagozlou, 2008. *in vitro* maturation of sheep oocytes in different concentrations of mare serum. *African J. Biotechnol.*, 7(18): 3380-3382.
- Noda, Y., H. Matsumoto, Y. Umaoka, K. J. Tatsumi, J. Kishi and T. Mori, 1991. Involvement of superoxide radicals in the mouse two-cell block. *Mol. Reprod. Dev.*, 28: 356-360.
- Ocaña Quero, J. M., M. Moreno Millán, M. Pinedo Merlin, M. A. Ortega Marisca A. Rodero Franganillo. 1999. *in vitro* bovine embryos production: influence of serum and hormonal supplementation: *Arch. Zootec.* 48: 71-74., *Archivos de zootecnia* vol. 48, núm. 181, p. 72.
- Parrish, J. J., J. L. Susko-Parrish and N. L. First, 1985. Effect of heparin and chondroitin sulfate on acrosome reaction and fertility of bovine sperm *in vitro*. *Theriogenology*, 24: 557-549.
- Poulin, N. Touzé, J. L. Pennetier, S. Schmaltz B. and Y. Cognié, 2006. *in vitro* production of ruminant embryos: results, limits and perspectives. 2006 symposium COA/INRA scientific cooperation in agriculture, Tainan (Taiwan, R. O. C.). pp: 59-78.
- Restall, B. J. and R. G. Wales, 1966. The fallopian tube of the sheep III. The chemical composition of the fluid from the fallopian tube. *Aust. J. Biol. Sci.*, 19: 687-698.
- Roa, B. S., K. S Naidu, D. Amarnath, R. Vagdevi, A. S. Roa, K. V. Brahmaiah and V. H. Roa, 2002. *in vitro* maturation of sheep oocytes in different media during breeding and non-breeding seasons. *Small Rumin. Res.*, 43: 31-36.
- Rosenkrans, C. F., G. Q. Zeng, G. T. McNamara, P. K. Schoff and N. L. First, 1993. Development of bovine embryos *in vitro* as affected by energy substrate. *Biol. Reprod.*, 49: 459-462.
- Samardžija M, Karadjole M, Cergolj M, Tomašković A, Dobranić T, Getz I, et al., The comparison of the two bull sperm separation methods for *in vitro* fertilization. *Tierarztl. Umschau.* 2005;60(4):193-199.
- Seré C, van de Zijpp A, Persley G and E Rege Animal Genetic Resources, *Information Bulletin* 2008; 42: 3-27.
- Shimada, M., Y. Yanai, T. Okazaki, N. Noma, I. Kawashima, T. Mori and J. S. Richards, 2008. Hyaluronan fragments generated by sperm-secreted hyaluronidase stimulate cytokine/chemokine production via the TLR2 and TLR4 pathway in cumulus cells of ovulated COCs, which may enhance fertilization. *Development*, 135: 2001-2011.
- Sirard, M. A., F. Richard, P. Blondin, C. Robert, 2006. Contribution of the oocyte to embryo quality. *Theriogenology*, 65(1): 126-136.
- Stringfellow, D. A. and S. M. Seidel, 1998. *Manual of the International Embryo Transfer Society: A Procedural Guide and General Information for the Use of Embryo Transfer Technology, Emphasizing Sanitary Precautions.* 3rd Edn., International Embryo Transfer Society, USA., ISBN: 9780966238600, Pages: 170.
- Suthar V. S. and Shah R. G. 2009: Bovine *In vitro* Embryo Production: review *Veterinary World*, vol. 2 (12) : 478-479. www.veterinaryworld.org
- Tamba, S., R. Yodoi, E. Segi-Nishida, A. Ichikawa, S. Narumiya and Y. Sugimoto, 2008. Timely interaction between prostaglandin and chemokine signaling is a prerequisite for successful fertilization. *Proc. Nat. Acad. Sci. USA*, 105: 14539-14544.
- Tanghe, S., A. Van Soom, H. Nauwynck, M. Coryn and A. De Kruif, 2002. Minireview: Functions of cumulus oophorus during oocyte maturation, ovulation and fertilization. *Mol. Reprod. Dev.*, 61: 414-424.
- Tervit, H. R., D. G. Whittingham and L. E. A. Rowson, 1972. Successful culture *in vitro* of sheep and cattle ova. *J. Reprod. Fertil.*, 30: 493-497.
- Thornton P K, Jones P G, Owiyo T, Kruska R L, Herrero P, Kristjanson A, Notenbaert H, Bekele N and A Omolo Report to the Biotechnology Department for International Livestock Research

- Institute (ILRI), 2006.
- Van Blerkam, J., H. Bell and D. Weipz, 1990. Cellular and developmental biological aspects of bovine meiotic maturation, fertilization and preimplantation embryogenesis *in vitro*. J. Electro. Microsc. Tech., 16: 298-323.
- Van Soom, A., S. Tanghe, I. De Pauw, D. Maes and A. DeKruif, 2002. Function of the cumulus oophorus before and during mammalian fertilization. Reprod Domest Anim, 37(3): 144-151.
- Watson, A. J., 2007. Oocyte cytoplasmic maturation: A key mediator of oocyte and embryo developmental competence. J. Anim. Sci., 85(E. Suppl.): E1-E3.
- Whitten, W. K., 1971. Nutritional requirements for the culture of preimplantation embryos *in vitro*. Adv. Biosci., 6: 129-139.
- Xu, K. P. and W. A. King, 1990. Effects of oviductal cells and heparin on bovine sperm capacitation *in vitro*. Biol. Reprod., 89-89.
- Yang, X., C. Kubota, H. Suzuki, M. Taneja, P. E. Bols and G. A. Presicce, 1998. Control of oocyte maturation in cows: Biological factors. Theriogenology, 49: 471-482.
- Yaumachi, N. and T. Nagai, 1999. Male pronuclear formation in denuded porcine oocytes after *in vitro* maturation in the presence of cysteamine. Biol. of Reprod. 61(3): 828
- Zhang, L., S. Jiang, P. J. Wozniak, X. Yang and R.A. Godke, 1995. Cumulus cell function during bovine oocyte maturation, fertilization and embryo development *in vitro*. Mol. Reprod. Dev., 40: 338 - 344.

How to cite this article:

Mosisa, D. B., 2022. Bovine *in vitro* embryo production and its contribution in improving animal products and productivity: A review. Int. J. Curr. Res. Biosci. Plant Biol., 9(2): 21-30.

doi: <https://doi.org/10.20546/ijcrbp.2022.902.004>